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UNIVERSITY

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FACULTY OF ENGINEERING & TECHNOLOGY
DEPARTMENT OF BIOTECHNOLOGY

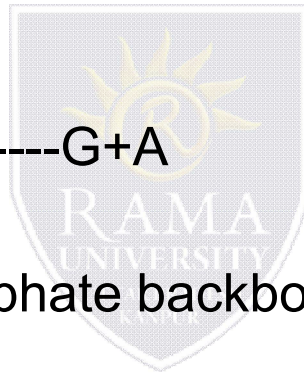
- Polynucleotide Kinase radioactive label at one 5' end of the DNA using gamma-³²P

5' G A C G T G C A A C G A A 3'

³²P **5' G A C G T G C A A C G A A 3'**

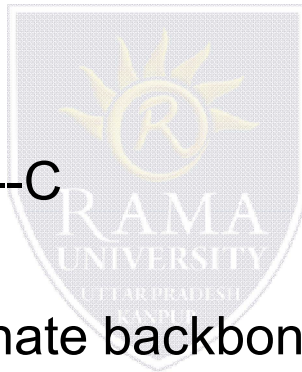
Chemical Modification and Cleavage

- Base Modification using Dimethyl sulphate
 - Purine
 - Adenine
 - Guanine
 - Only DMS----- G
 - DMS+ Formic acid-----G+A
- Cleavage of Sugar Phosphate backbone using Piperidine

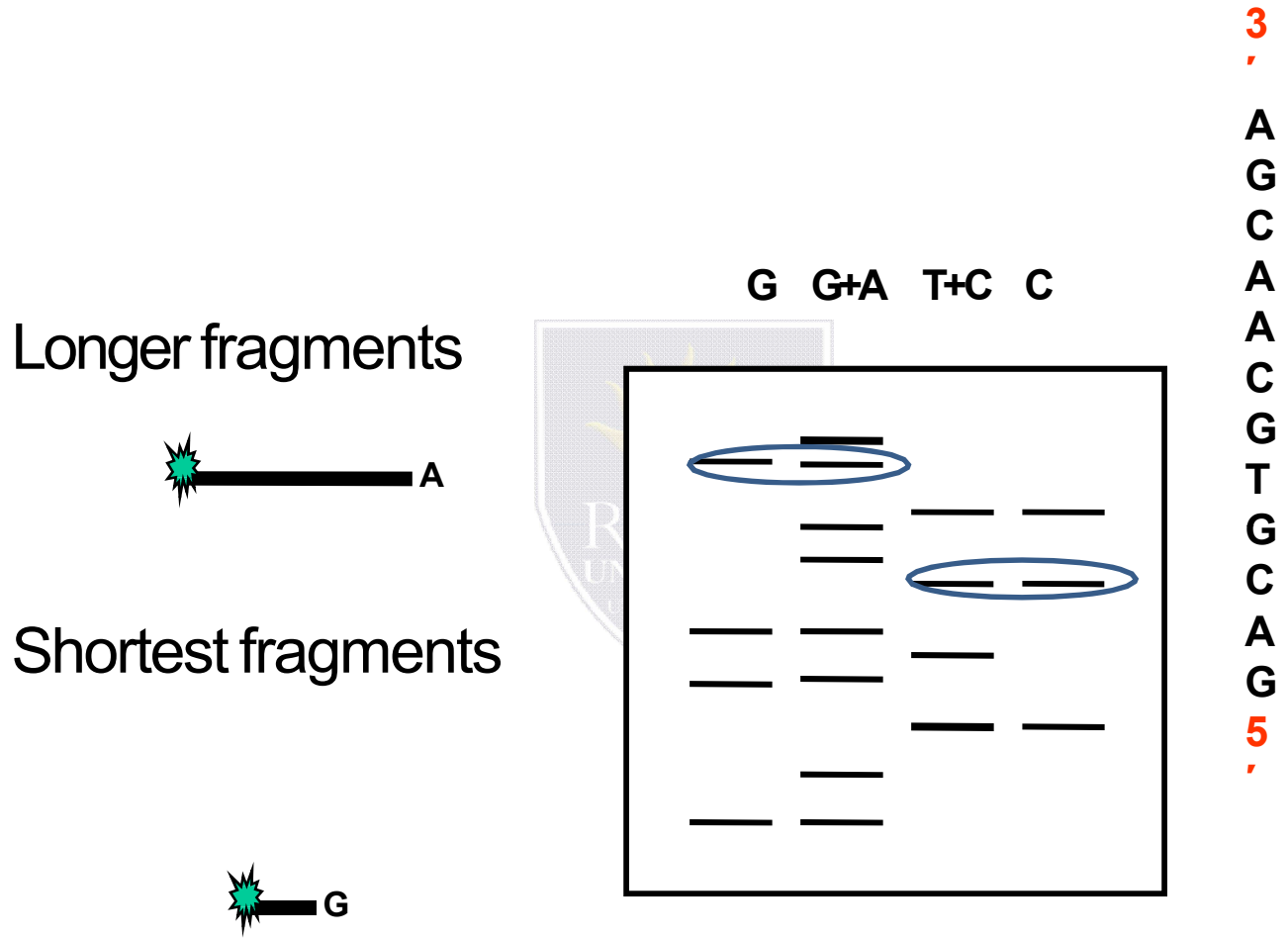


Chemical Modification and Cleavage

- Base modification using Hydrazine
 - Pyrimidine
 - Cytocine
 - Thymidine
 - Hydrazine----- C+T
 - Hydrazine + NaCl-----C
- Cleavage of Sugar Phosphate backbone using Piperidine



Maxam-Gilbert Sequencing

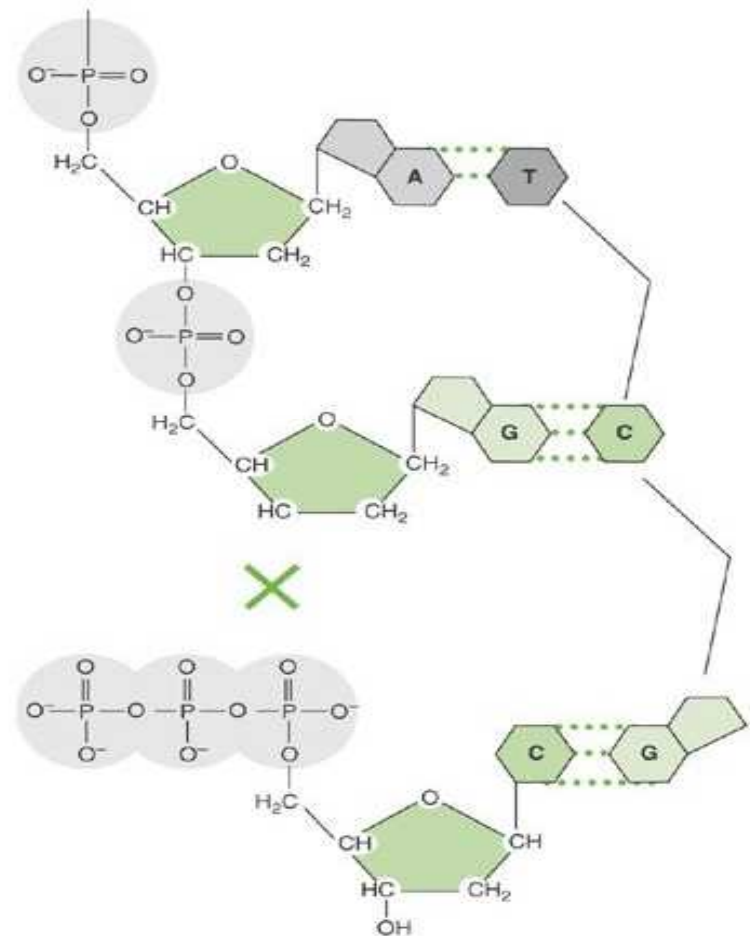
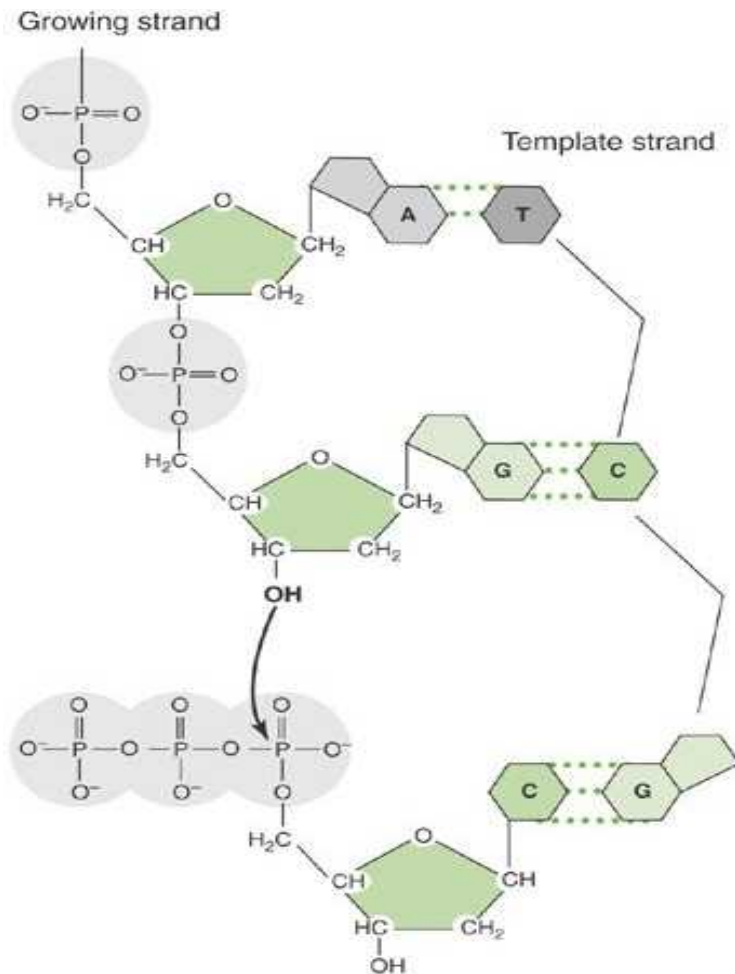


Sequencing gels are read from **bottom to top** (5' to 3').

³²P **5'** G A C G T G C A A C G A

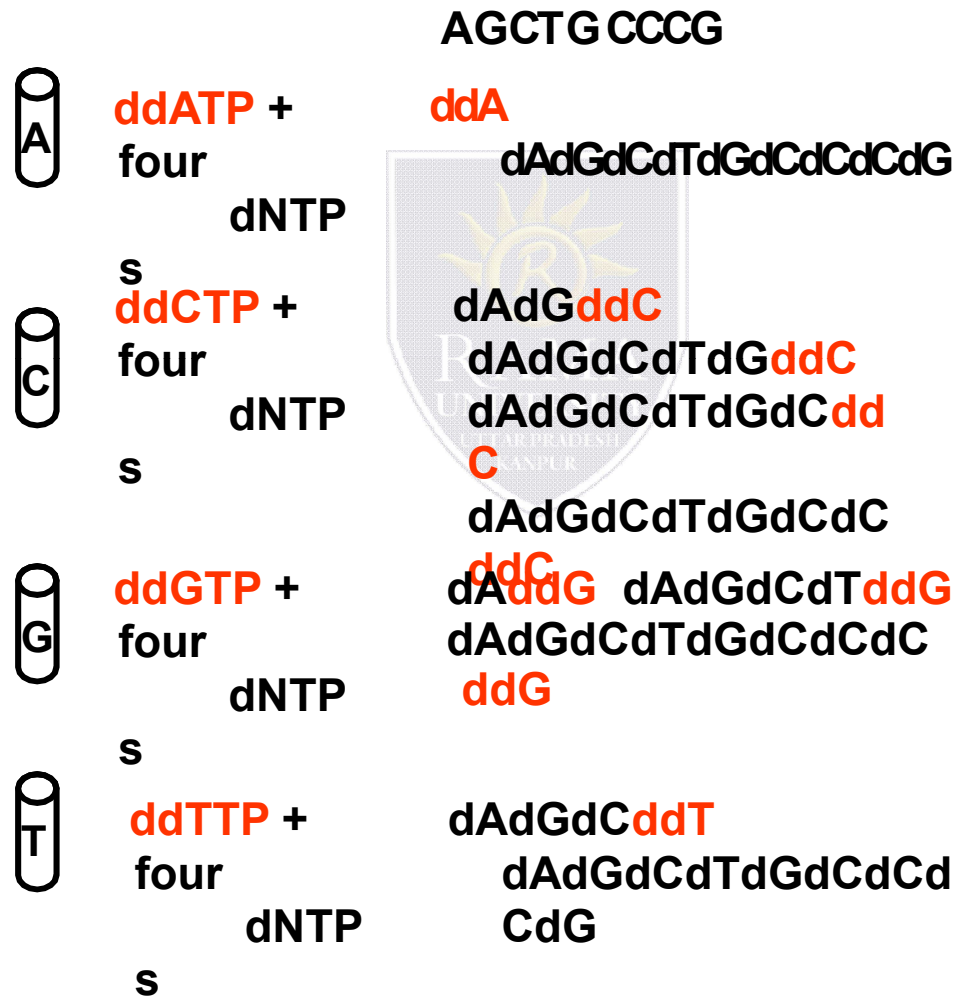
Maxam Gilbert Sequencing: Process Summarized

1. Label 5'- end of DNA
 2. Aliquot DNA sample in 4 tubes
 3. Perform base modification reaction
 4. Perform Cleavage reaction
 5. Perform Gel Electrophoresis
 6. Perform Autoradiography
 7. Interpret results
-



The 3'-OH group necessary for formation of the phosphodiester bond is missing in ddNTPs

Sanger, Chain Termination Sequencing

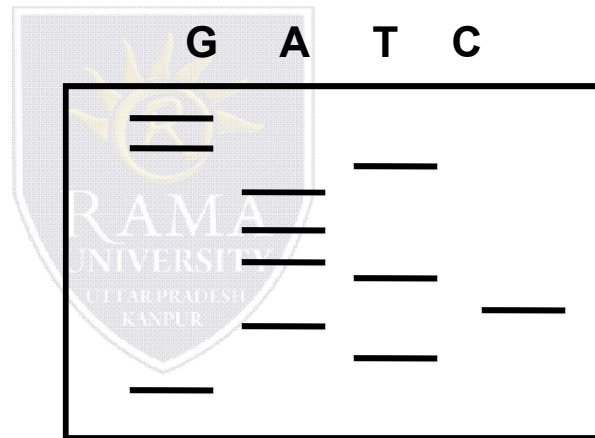


Chain Termination Sequencing

Longer fragments



Shorter fragments



3
,
G
G
T
A
A
A
T
C
A
T
G
5
,

Sequencing gels are read from **bottom to top** (5' to 3')

5'-TACACGATCGA-3'

3'-ATGTGCTAGCT-5'

Denature the sequence

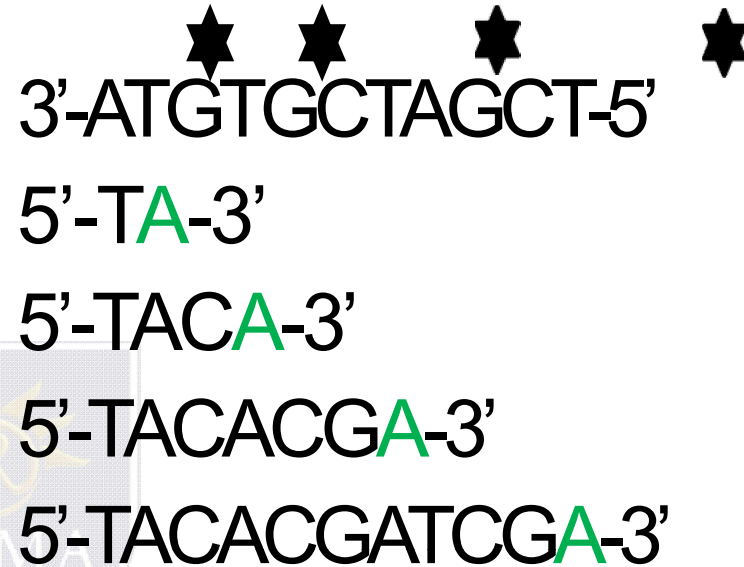
Use only forward primer i.e. using 3'-5'



Amplification in ddTTP



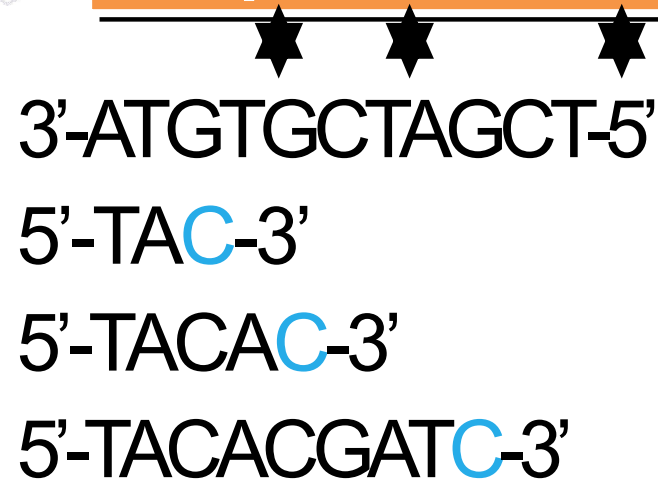
Amplification in ddATP



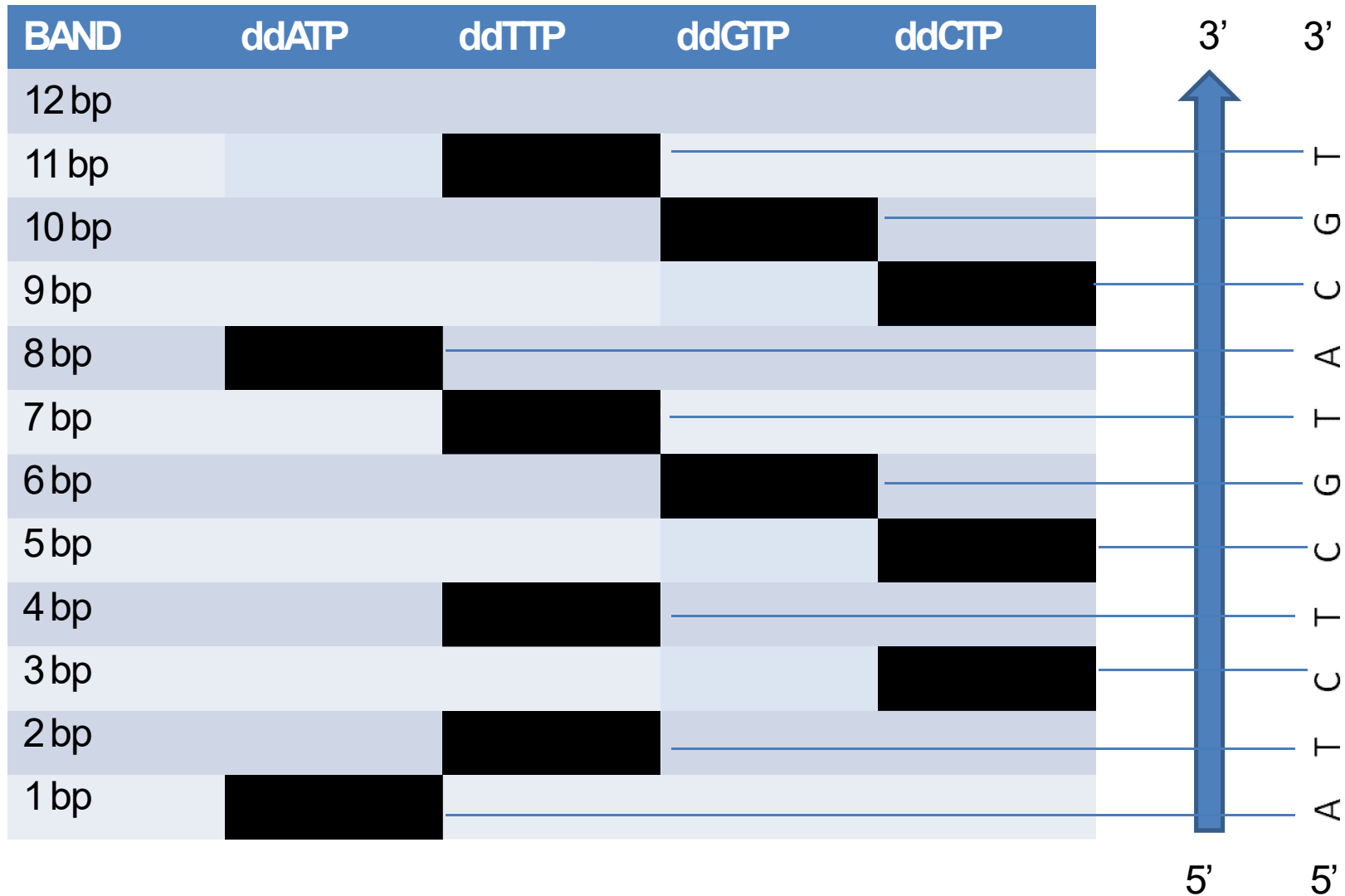
Amplification in dGTP



Amplification in ddCTP



Reading Sequence



Sanger Sequencing: Process Summarized

1. Get enough quantity of DNA (Run PCR)
 2. Aliquot DNA into four different tubes
 3. Prepare PCR reaction mix as below:
 - Primer, taq PM, template(ss DNA), dNTPS (All) and ddNTPs(ddATP, ddGTP,ddCTP & ddTTP respectively)
 4. Run PCR
 5. Perform Gel Electrophoresis
 6. Interpret results
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