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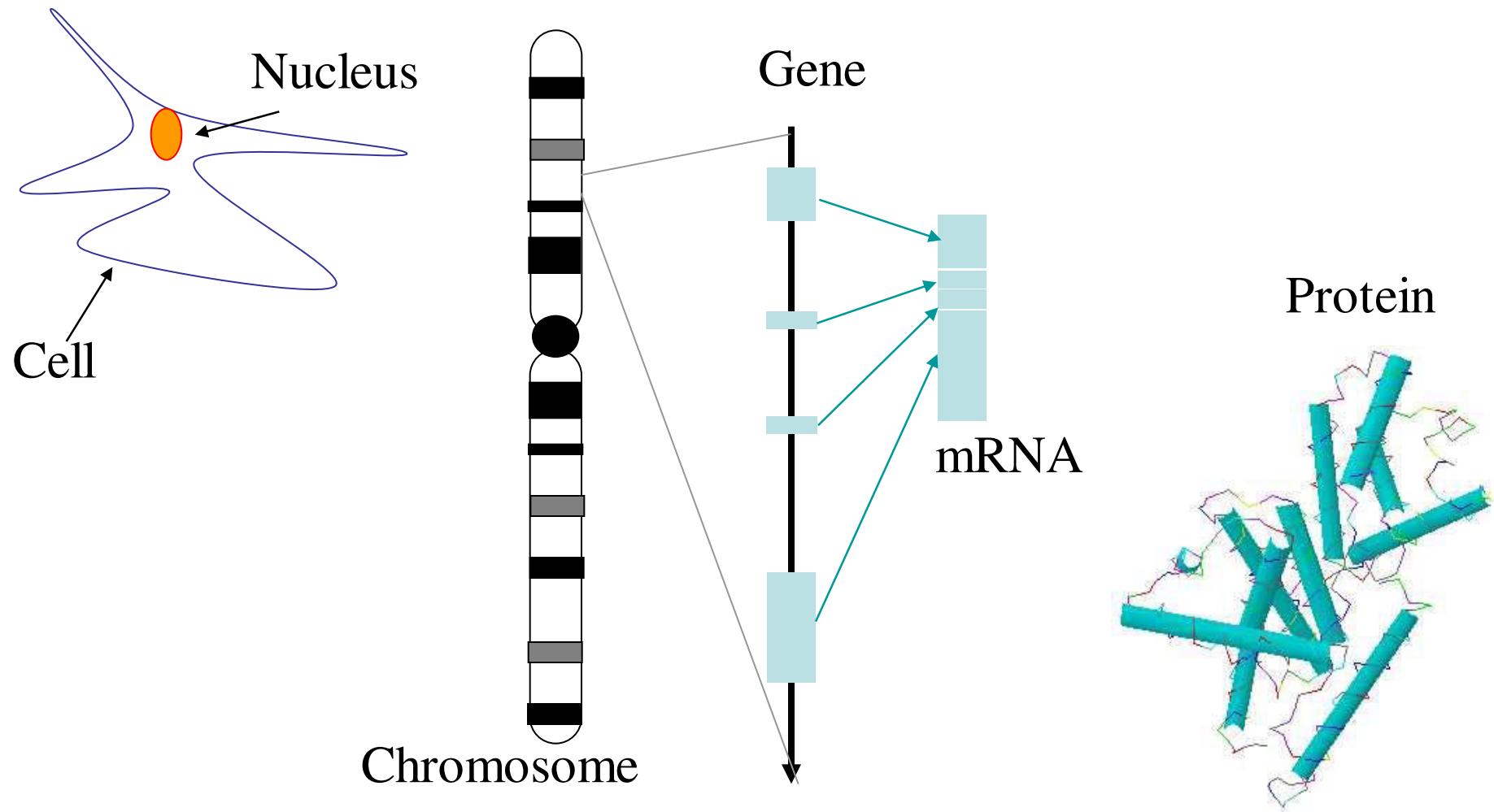
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FACULTY OF ENGINEERING & TECHNOLOGY DEPARTMENT OF BIOTECHNOLOGY

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Genome Annotation

Cells, Chromosomes, DNA, and Genes



Definitions

- Unless otherwise stated, annotation refers to prediction of protein-coding genes
- Methods exist to annotate
 - tRNA, rRNA
 - Several other small RNAs
 - Repetitive elements
 - [microRNAs]

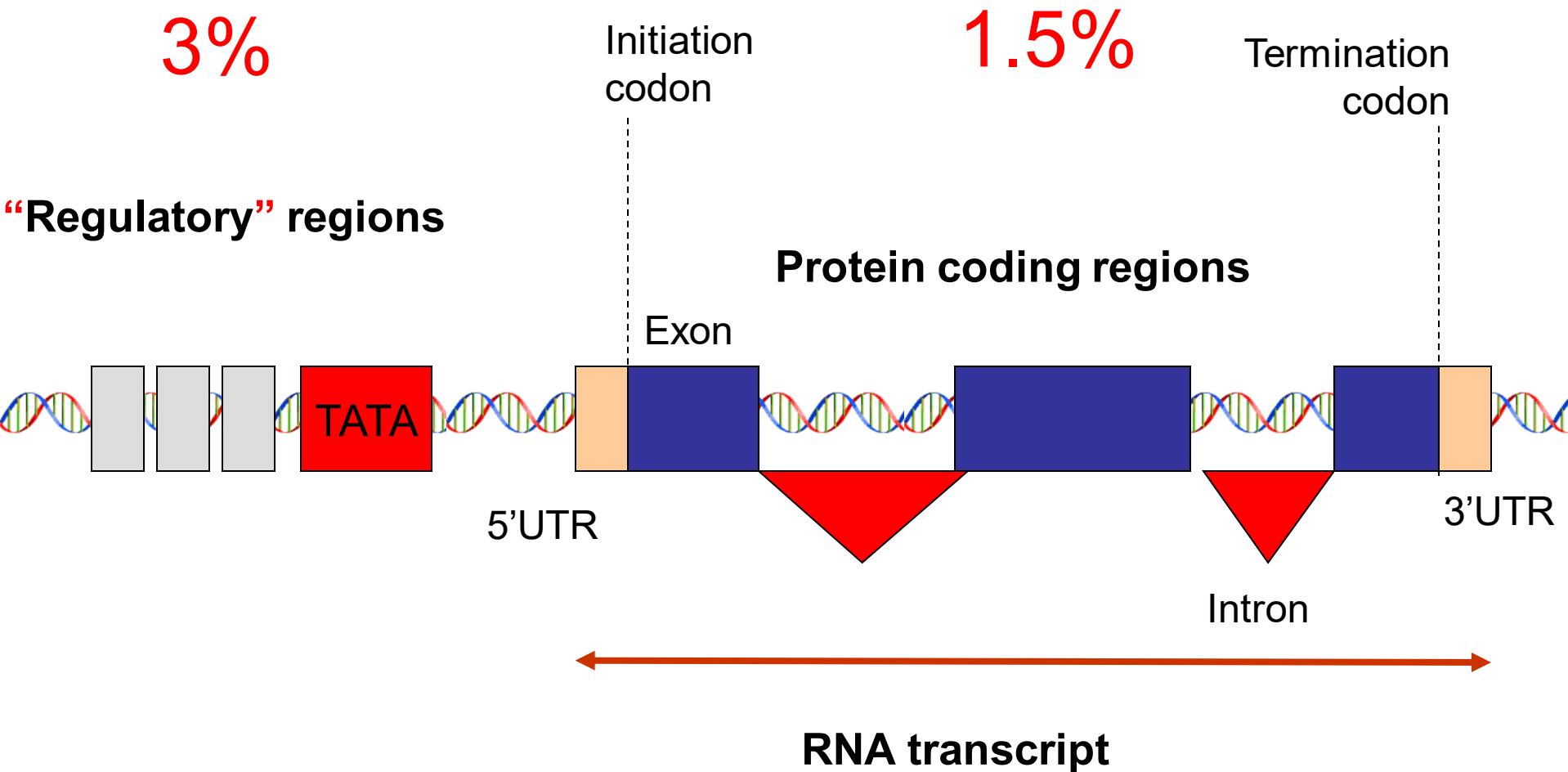
Challenges

- Signal to noise
 - 1% protein coding sequence
 - Pseudogenes
- Splicing
 - Discontinuous nature of eukaryotic genes
 - Alternative splicing
- Non-uniform genome characteristics
 - Range of G+C content
 - Range of mutation rates
 - Gene/Genome segment duplication

1% codes for protein

3% conserved non-coding

Structure of Genes

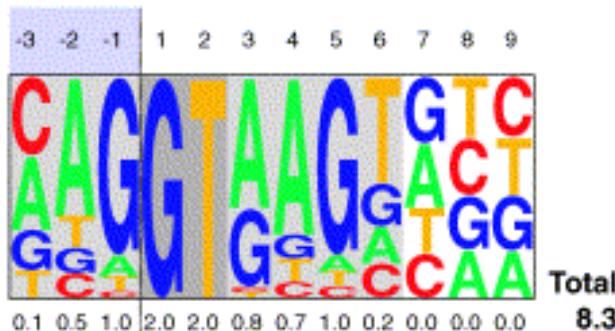


Genome Annotation Methods

- Known Genes
 - Blastn cDNA against genome
- Protein similarity
 - Blastx genome against SWISS-PROT
- Genome-Genome alignment
 - Blastz
- *De novo* prediction
 - GRAIL, Genscan, FGENESH
- Integrated methods
 - Expert review, ‘Combiner’ algorithms

Signal Detection: Splice Sites

Donor

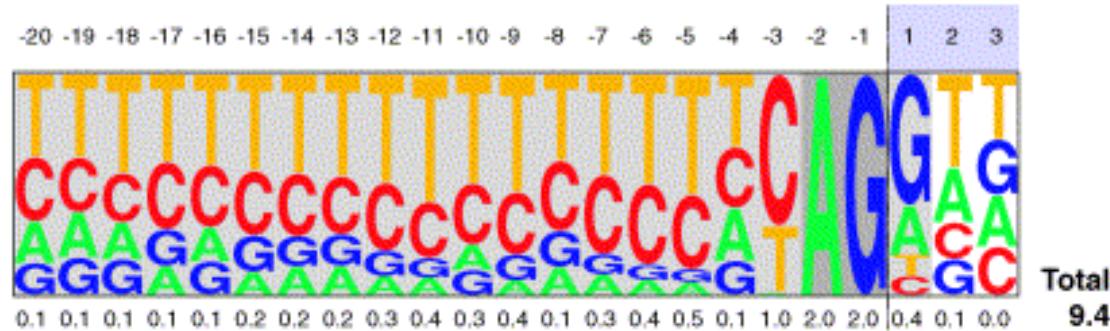
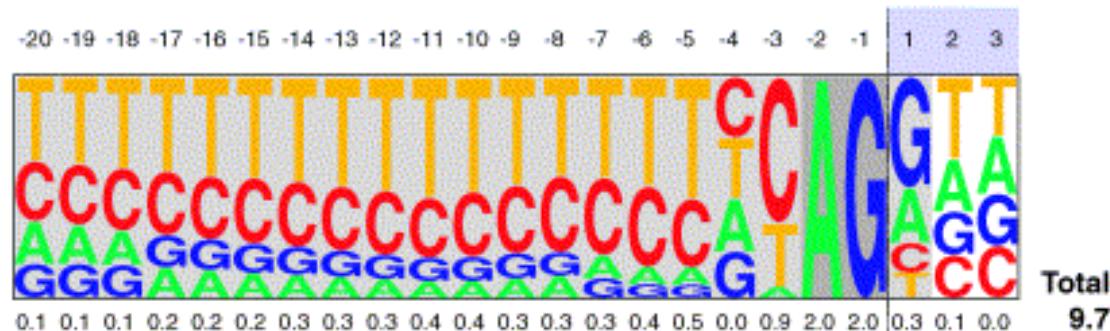


Total 12.6

Sequence logo showing the sequence ACTGAACTTCAAGGGATGTATTCCTTTCCG. The x-axis shows positions -3, -2, -1, 1, 2, 3, 4, 5, 6, 7, 8, 9. The y-axis shows probabilities for A, C, G, T.

Position	A	C	G	T
-3	0.0	0.0	0.1	1.1
-2	0.0	0.0	0.1	2.0
-1	0.0	0.0	0.1	2.0
1	0.0	0.0	0.1	2.0
2	0.0	0.0	0.1	2.0
3	0.0	0.0	0.1	2.0
4	0.0	0.0	0.1	2.0
5	0.0	0.0	0.1	2.0
6	0.0	0.0	0.1	2.0
7	0.0	0.0	0.1	2.0
8	0.0	0.0	0.1	2.0
9	0.0	0.0	0.1	2.0
Total	0.0	0.0	0.5	14.9

Acceptor

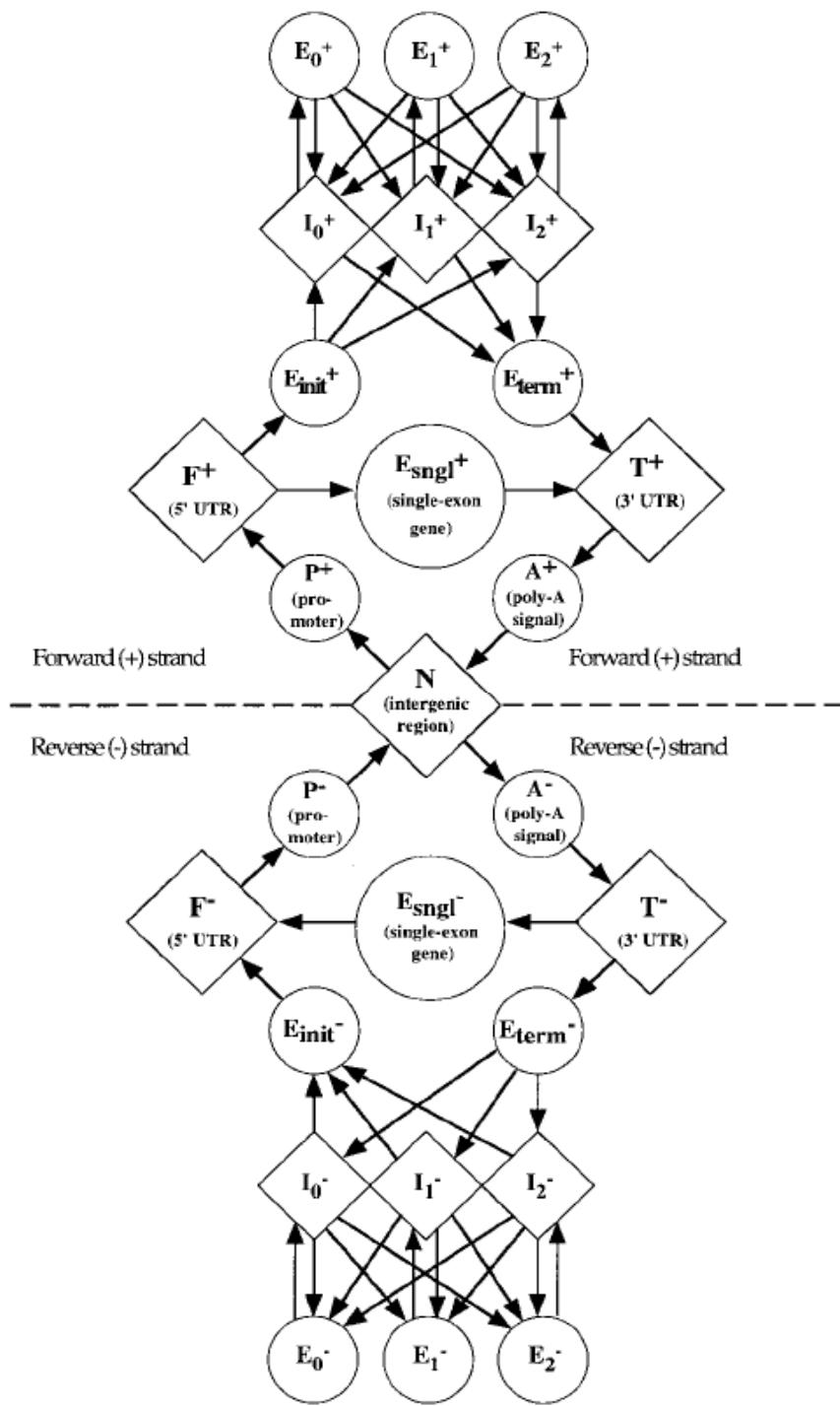


Branch point



A sequence logo representing the sequence CTCCTAGGAAACCCAGTGTGCCCTAGAATAGCTT. The y-axis indicates positions from -6 to 3. The x-axis shows the four bases: A (green), T (blue), C (red), and G (yellow). The logo shows a strong bias towards 'C' at positions -6, -5, -4, -3, and -2, and towards 'A' at positions 1, 2, and 3.

Genscan's View of a Gene



E = Exon

I = Intron

A = polyadenylation signal

P = Promoter

F, T = UTR

N = Intergenic sequence

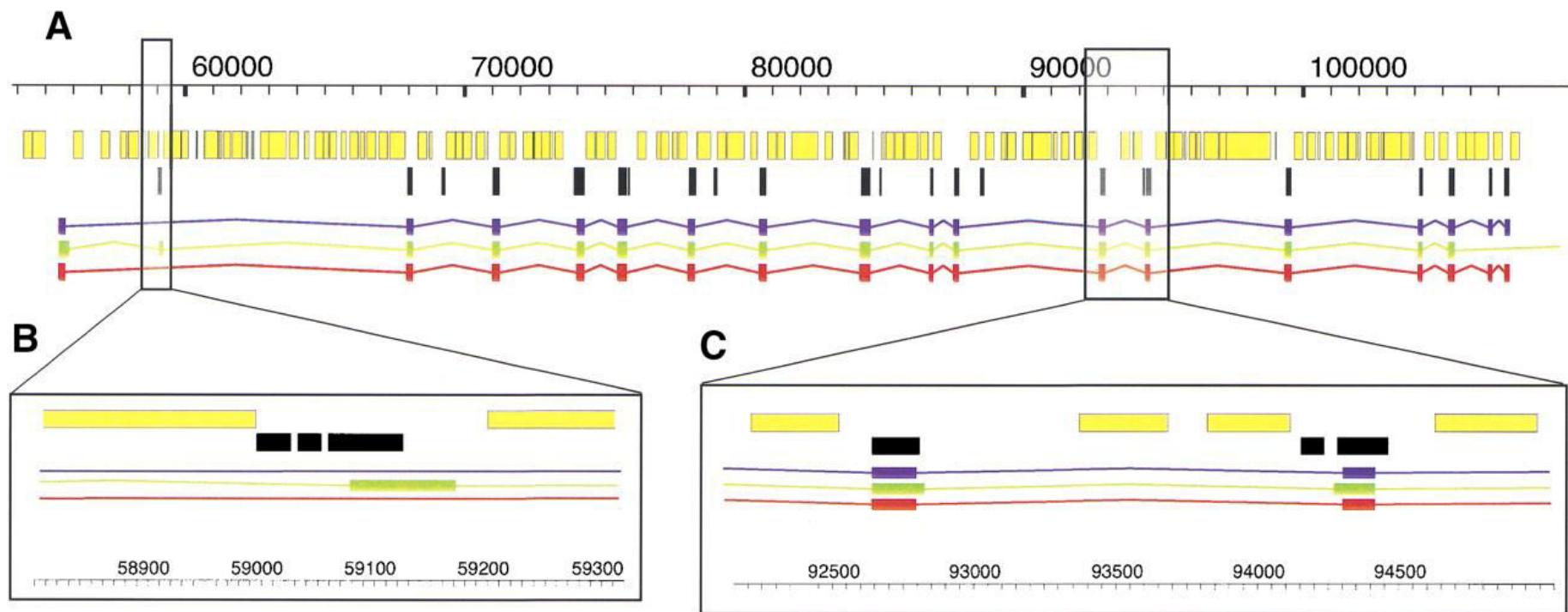
De novo methods: Single genome

- GRAIL
 - Uberbacher & Mural *PNAS* 88:11261, 1991
 - Neural network
 - Trained using known gene structures
- GENSCAN
 - Burge & Karlin *J. Mol. Biol.* 268:78, 1997
 - Generalized hidden Markov model approach
 - Probabilistic model of gene structure
 - Uses descriptions of transcriptional, translational, and splicing signals
 - Distinct parameter sets for varying gene density and structure across G+C ranges
 - Allows for partial genes, multiple genes, and genes on both strands

De novo methods: Dual genome

- Pair HMM approach (SLAM)
 - Joint probability model for sequence alignment and gene structure definition
 - Dynamic programming algorithm combines classic alignment algorithms and HMM decoding
- ‘Informant genome’ approach (SGP-2, TWINSCAN)
 - Alignments performed first (BLASTN, TBLASTX)
 - Alignments ‘inform’ prediction algorithms based on single-genome predictors (e.g. GENSCAN)

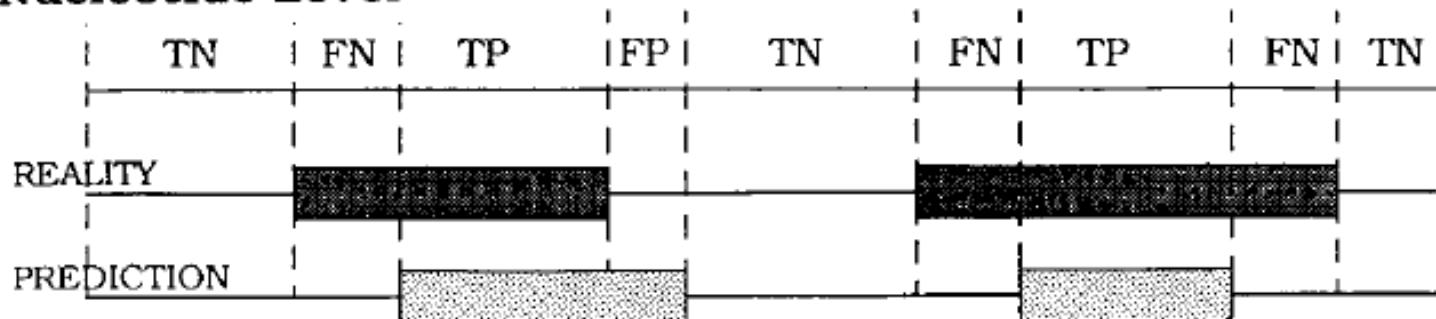
Genscan vs Twinscan



A detailed view of a TWINSCAN prediction (red), a GENSCAN prediction (green), and an aligned RefSeq transcript (blue). Masked repetitive and low-complexity regions (yellow) and mouse alignments (black) are indicated. (A) Complete gene prediction at the *KIAA1630* gene (NM_018706) from *Homo sapiens* 10p14. Note that the presence of conservation is neither a necessary (e.g., the first exon), nor a sufficient (e.g., the first alignment block condition) for TWINSCAN to predict an exon. (B) A magnified region around the second exon predicted by GENSCAN. TWINSCAN correctly omits this exon because the conserved region ends within it. (C) A magnified region around the 11th and 12th RefSeq exons. TWINSCAN correctly predicts both splice sites because they are within the aligned regions.

Evaluation of Predictions

Nucleotide Level



		REALITY		
		coding	no coding	
PREDICTION	coding	TP	FP	TP+FP
	no coding	FN	TN	FN+TN
		TP+FN	TF+TN	

$$S_n = \frac{TP}{TP + FN}$$

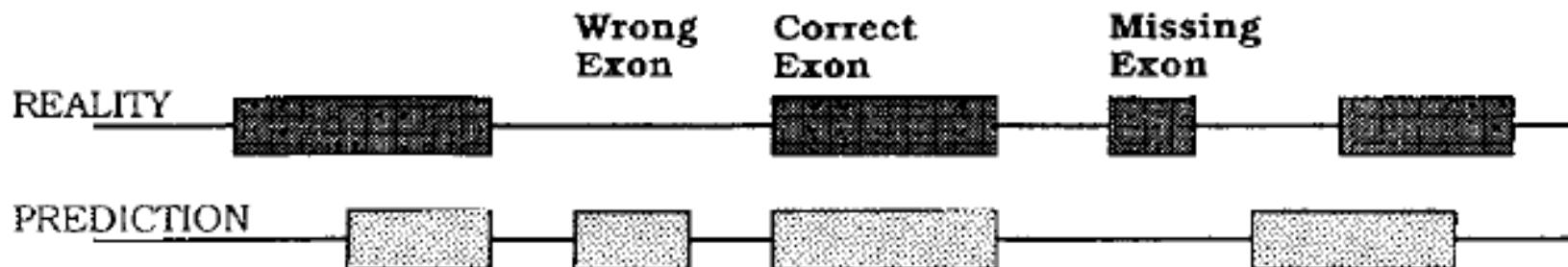
Sensitivity

$$Sp = \frac{TP}{TP + FP}$$

Specificity

Evaluation of Predictions

Exon Level



$$Sn = \frac{\text{number of Correct Exons}}{\text{number of Actual Exons}}$$

Sensitivity

$$Sn = \frac{\text{number of Correct Exons}}{\text{number of Predicted Exons}}$$

Specificity

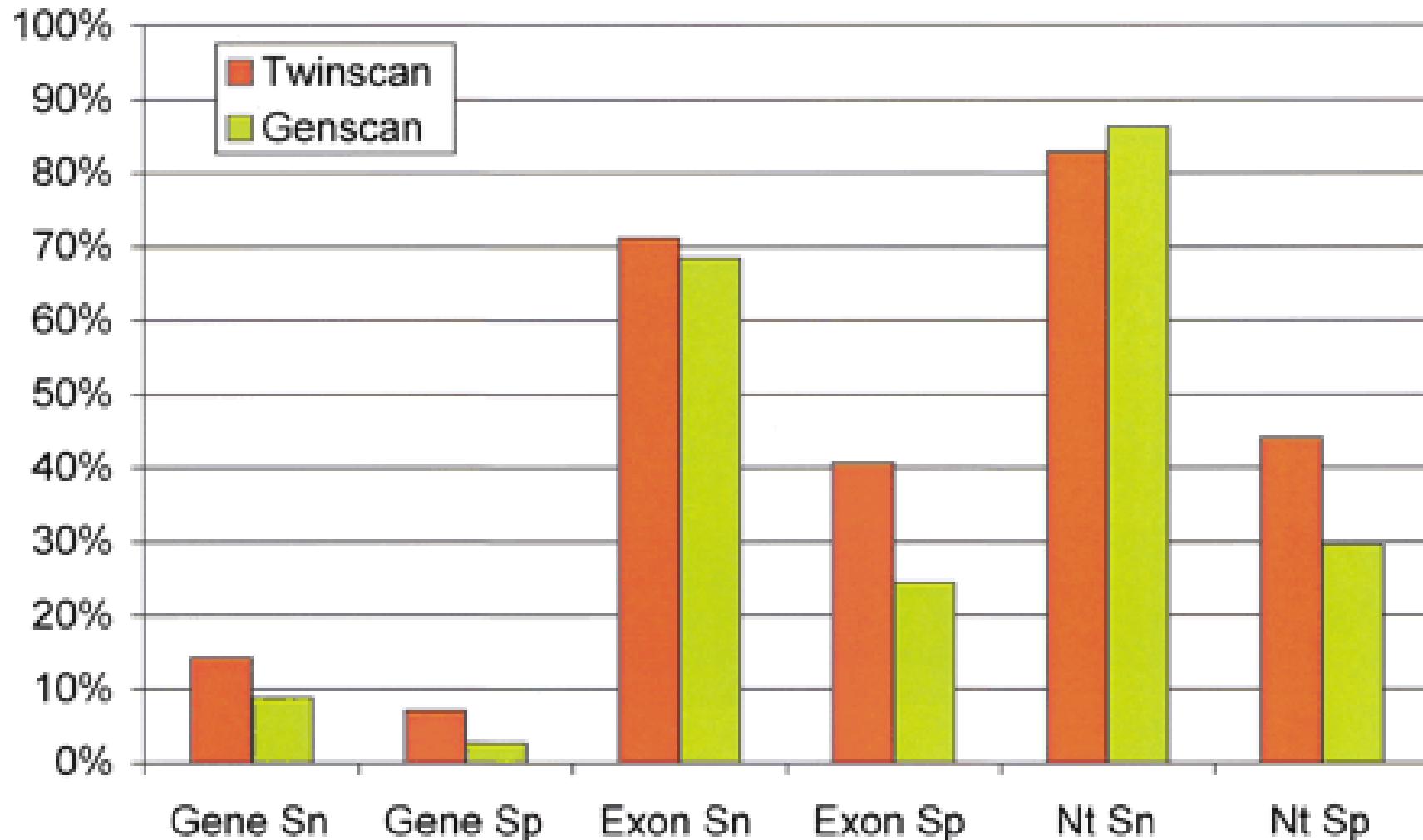
$$ME = \frac{\text{number of Missing Exons}}{\text{number of Actual Exons}}$$

(**Sensitivity**)

$$WE = \frac{\text{number of Wrong Exons}}{\text{number of Predicted Exons}}$$

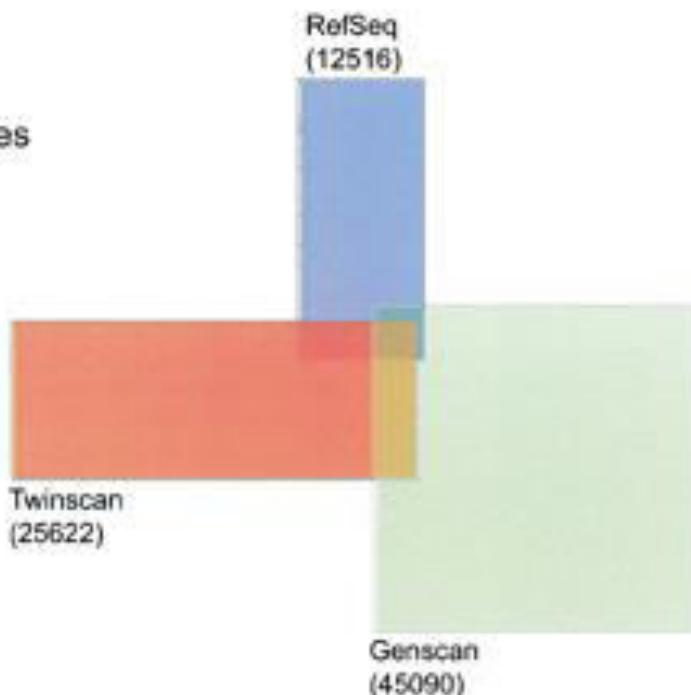
(**Specificity**)

Annotation of the Mouse Genome



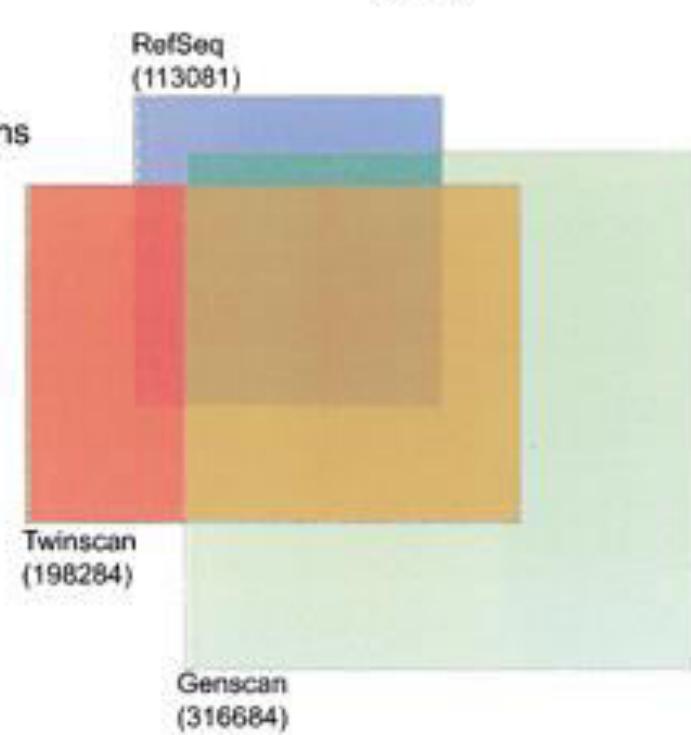
A

Exact Genes



B

Exact Exons



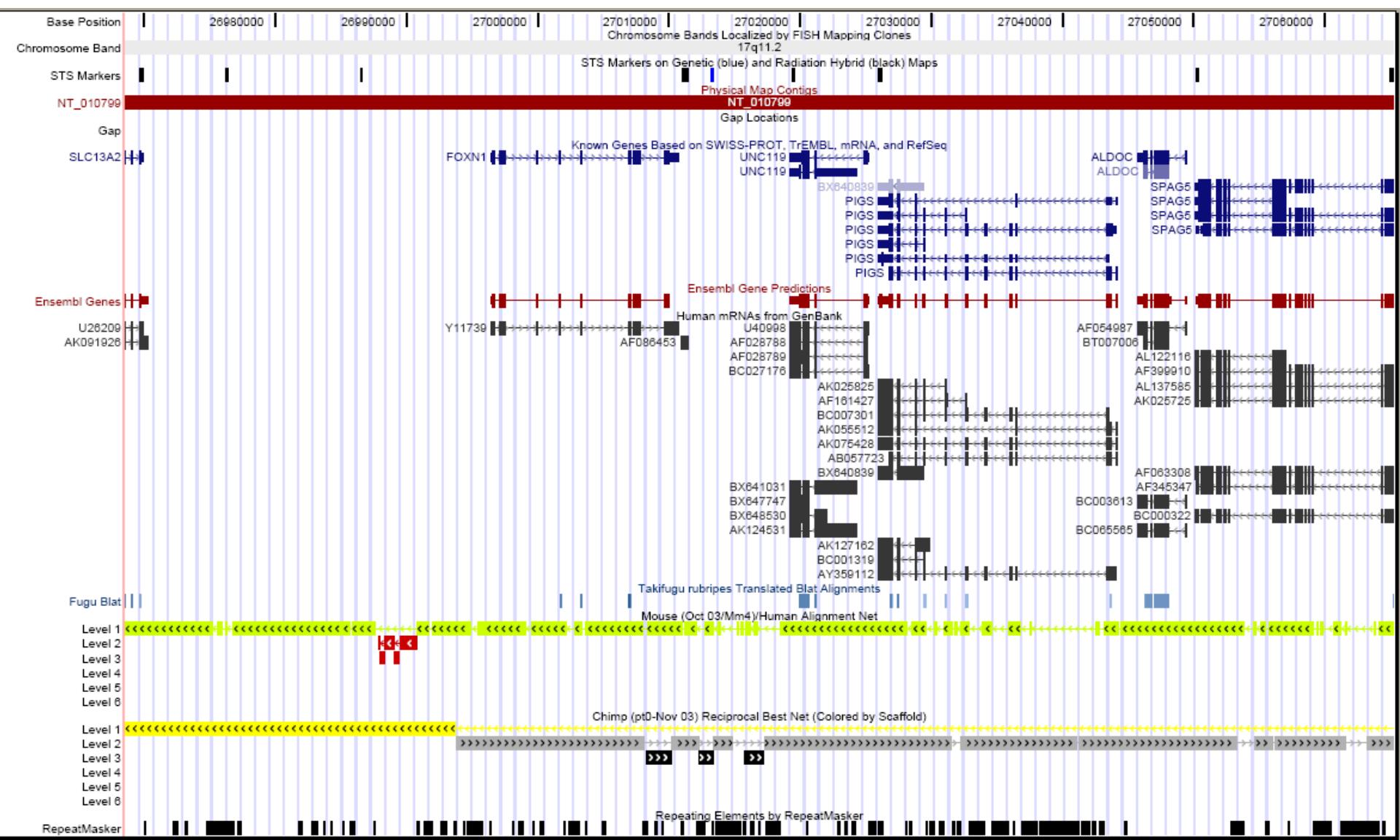
Assessment of Genscan and Twinscan

Addition of Evidence

- Known cDNAs
- ESTs (partial cDNA sequence)
- Known genes
- [Predicted genes from other species]
- Genome comparison
- Repeat-masking



Genome Browser



Additional Reading

- Brent & Guigo, Recent advances in gene structure prediction.
Curr. Op. Struct. Biol. 14(3) 264-272, 2004
- Fickett, JW. The gene identification problem: an overview for developers.
Comput. Chem. 20:103-118, 1996